

CONSTITUENTS OF *TABEBUIA GUAYACAN* THE STRUCTURE OF GUAYACANIN

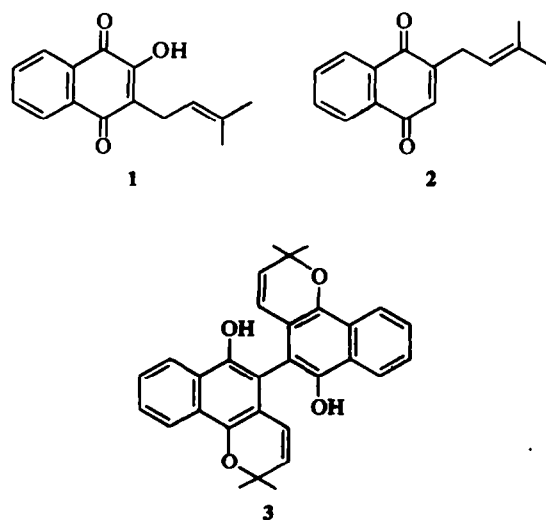
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Abstract—The heartwood of *T. guayacan* has yielded tectol and a new, novel dibenz-xanthene derivative (guayacandin). The structure of guayacandin suggests its biogenetic derivation through the condensation of prenylnaphthalene quinols. The tetrahydro derivative of the xanthene was synthesized by an analogous acid catalyzed reaction of prenylnaphthalene quinol derivatives of lapachol and 1,4 naphthalene diol.

Long term marine exposure tests¹ have indicated the heartwood of the Central American tree, *Tabebuia guayacan* Hemsl., to be resistant to attack by marine boring organisms. At the invitation of the U.S. Naval Research Laboratory, an examination of the heartwood extractives of this tree has been undertaken in an effort to determine the identity of the natural protective constituent(s). With the exception of an early report indicating the presence of lapachol (1),² the constituents of *T. guayacan* have not previously been investigated. However, related *Tabebuia* species (*T. chrysantha* and *T. avellanedae*) have been examined by Thomson^{3,4} and others,^{5,6} leading to the identification of lapachol (1), several lapachol related naphthaquinones, and anthraquinones in this genus. The presence of deoxylapachol (2) has also been noted in an unidentified *Tabebuia* species.⁷ Lapachol and several of these known naphthaquinone derivatives have been detected in the present investigation of *Tabebuia guayacan*. In addition, tectol (3), a colorless dimer previously obtained from teak wood (*Tectona* sp.),⁸ and a new dimeric compound (guayacandin) have now been isolated and structurally identified.

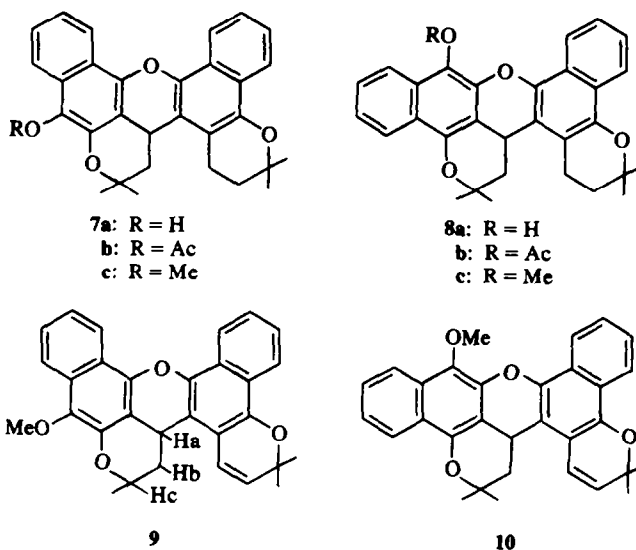
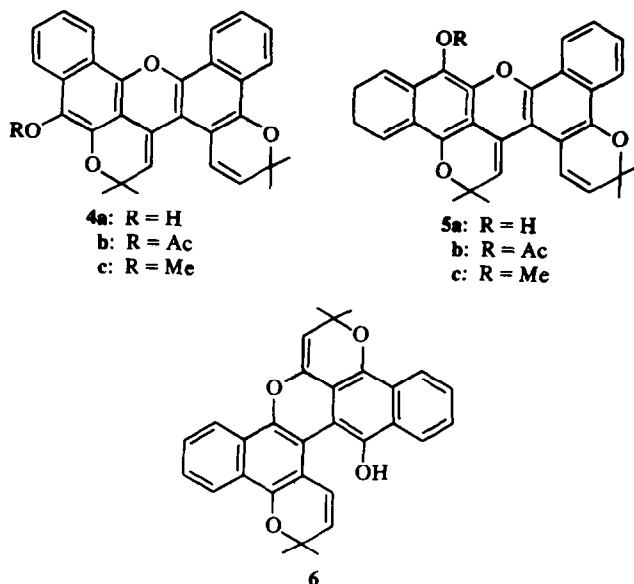
Ether and acetone extracts of the wood gave a colorless dimer, C₃₀H₂₆O₄ (m.p. 212°), and a slightly yellow compound, C₃₀H₂₄O₄ (m.p. 225–226°). The colorless dimer contains two phenolic OH groups and forms diacetyl,



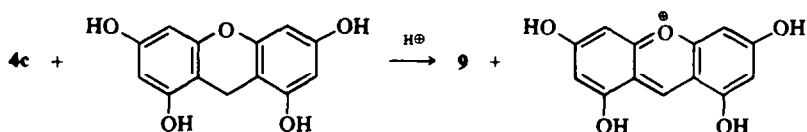
dimethyl, and tetrahydro derivatives, whose m.ps closely correspond with those reported for tectol diacetate, di-O-methyl-tectol and tetrahydrotectol. Although the NMR spectrum of tectol(3) does not appear to have been reported, the NMR spectrum of the *T. guayacan* product (Experimental) is in complete accord with the tectol structure 3.

The yellow compound, C₃₀H₂₄O₄, for which we propose the name *guayacandin*, forms a monoacetate (m.p. 248–249°) and a monomethyl ether (m.p. 205°) and, therefore, contains a single phenolic OH group. Guayacandin yields immediately a characteristic deep purple pyrylium salt (λ_{\max} 550 nm) on addition of HCl to its solution in ethanol (λ_{\max} 405 nm). The 100 MHz NMR spectrum of guayacandin in CDCl₃ shows two chromene gem dimethyl groups as 6H singlets at δ 1.56 and δ 1.58, an aromatic OH as a broad singlet at δ 5.60 and two vicinal vinylic (chromene) protons as doublets ($J = 10$ Hz) at δ 5.73 and δ 7.05. A non-coupled vinylic proton signal is at δ 5.80 and eight aromatic protons are in three multiplet resonances at δ 7.42–7.70 (4H), δ 8.00–8.35 (2H) and δ 8.36–8.65 (2H). These data are consistent with three possible structures, viz. 4a, 5a, or 6, each containing a chromene ring and a benzylically substituted chromene ring. Guayacandin can be further formulated as 4a or 5a rather than 6, on the basis of its catalytic hydrogenation. Compound 6 would be expected to consume three moles of hydrogen with benzylic ether hydrogenolysis to form tetrahydrotectol. Compounds 4a or 5a, on the other hand, should take up only two moles of hydrogen to form tetrahydroguayacandin 7a or 8a. Guayacandin, in fact, takes up only 2 moles of hydrogen to form the tetrahydro derivative. In accord with structure 7a or 8a, tetrahydroguayacandin in acid media undergoes facile oxidation to a compound which in turn forms a deep purple pyrylium salt with acids. This color reaction is characteristic of the conversion of xanthenes of xanthylum salts,^{9,10} and this compound must therefore be isomeric with the parent phenol of 9 or 10, and have a xanthene rather than a chromene double bond.

Inspection of the guayacandin structure (4a or 5a) indicates the benzylically substituted chromene ring should be much more readily protonated (with formation of the stable tertiary carbonium ion) than the second, unsubstituted chromene ring. This observation was used to prepare a crystalline dihydro derivative, 9 or 10, of guayacandin by selective reduction via an acid-catalyzed hydride ion transfer reaction between O-



methylguayacanine and 1,3,6,8-tetrahydroxy xanthene: e.g.,



The coupling of the benzylic H, proton with geminally coupled allylic protons, shown in Fig. 1, confirms structure 9 or 10 for this dihydro derivative, and, therefore, structure 4a or 5a for guayacanine.

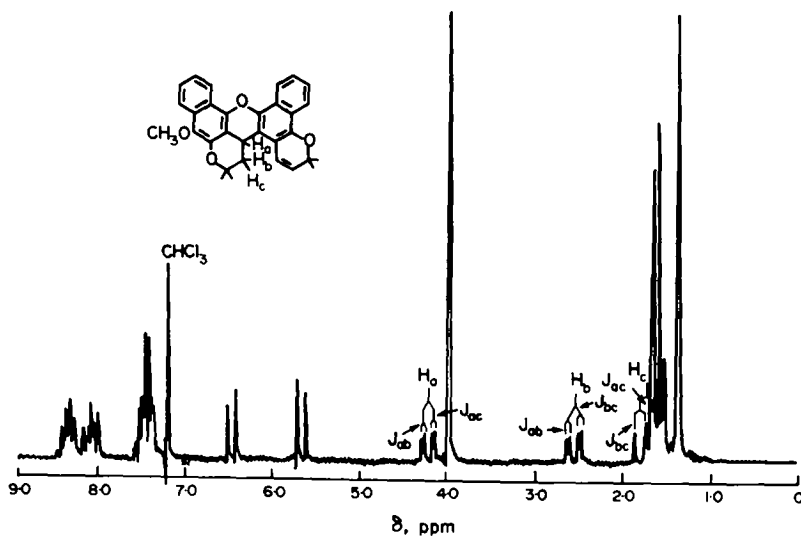
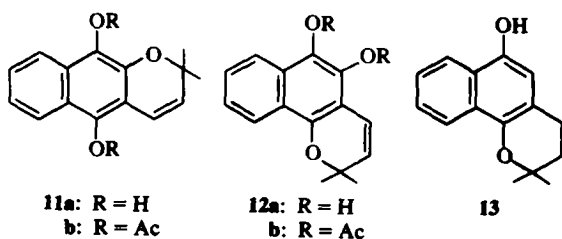
Synthesis of tetrahydroguayacanine

The inability to specifically formulate guayacanine as either 4a or 5a on the basis of its chemical and spectral properties, prompted an attempt to synthesize the two tetrahydro-structures 7a and 8a, one of which should then be identical with the hydrogenated natural product. Consideration of the two possible guayacanine structures suggests the molecule is biosynthetically derived by a condensation of the chromenes derived from the oxida-

tive cyclization of the quinols corresponding to lapachol (1) and deoxylapachol (2). On this basis, a biogenetic type synthesis of tetrahydroguayacanine was devised which would involve an acid catalyzed condensation of dehydro- α or β -lapachone diol (11a and 12a), from lapachol,¹¹ with nordihydrolapachenole (13) (Scheme 1).

As models for this synthetic approach, it was shown that 11a and 12a condensed readily with naphthalene-1,4 diol in aqueous formic acid to give, after acetylation, the analogous crystalline diacetates 14 and 15.

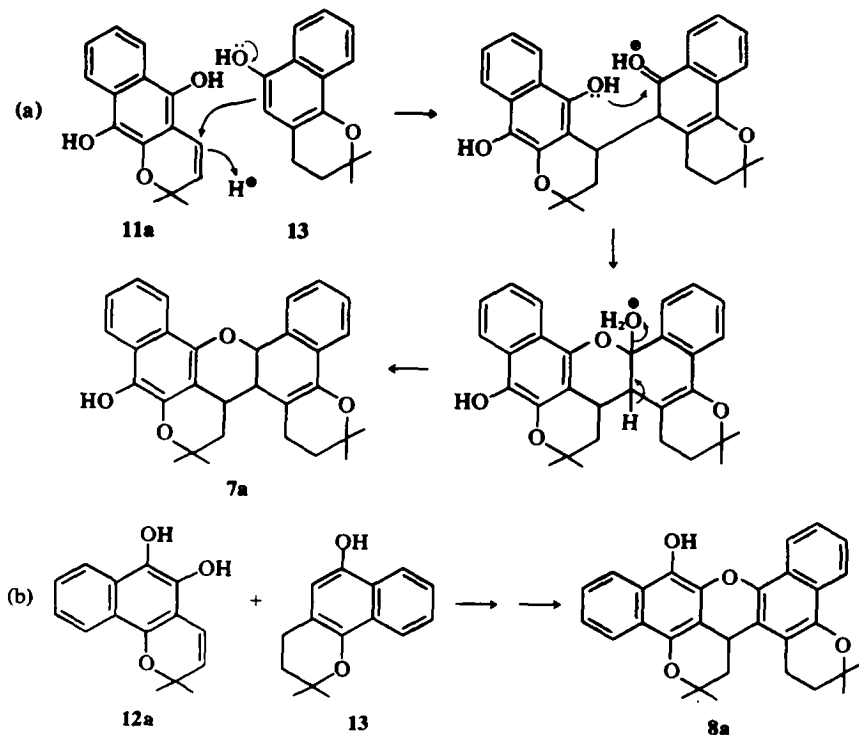
Extension of these model condensations to the synthesis of tetrahydroguayacanine required a convenient source of nordihydrolapachenole 13. This had been previously prepared in low yield (6%) by the Friedel-


 Fig. 1. The 100 MHz NMR spectrum of O-methyl dihydroguayacatin in CDCl_3 .


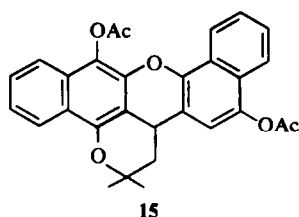
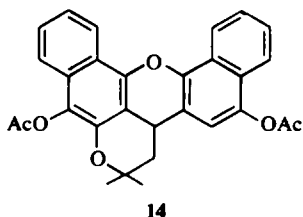
reaction involving a condensation (under N_2) of naphthalene-1,4-diol with 3-methylbut-2-ene-3-ol in aqueous formic acid containing ascorbic acid. Nordihyrolapachenole **13** readily condensed with **11a** in aqueous formic acid (under N_2) to yield a colorless crystalline compound (43% yield) which was identical in all respects with tetrahydroguayacatin from the natural source. The analogous reaction of **12a** with **13** gave a colorless crystalline compound (8% yield) which, surprisingly, also proved to be identical with the naturally derived tetrahydroguayacatin.

Crafts reaction of 1-bromo-3-methylbut-2-ene with naphthalene-1,4-diol⁴ and by the reductive cyclization of synthetic deoxylapachol⁴ (14.7% overall yield). Better yields (19%) of **13** were now obtained in a one-step

Thus, although tetrahydroguayacatin has been successfully synthesized, it is clear that a rearrangement of a condensed intermediate of dehydro α - or β -lapachone



Scheme 1.



diol, as observed by Ettlinger¹² for α - and β -lapachone in aqueous acids, occurs in either the condensation of 13 with 11a or of 13 with 12a and, therefore, that the synthesis does not unambiguously establish guayacanic acid as 4a or 5a.†

The structure of guayacanic acid was unequivocally established as 4a by X-ray crystallographic analysis. The X-ray molecular structure determination‡ is illustrated in the ORTEP¹³ drawing shown in Fig. 2. There are four possible conformations of the molecule defined by the orientation of the two *gem* dimethyl groups of the chromene rings.

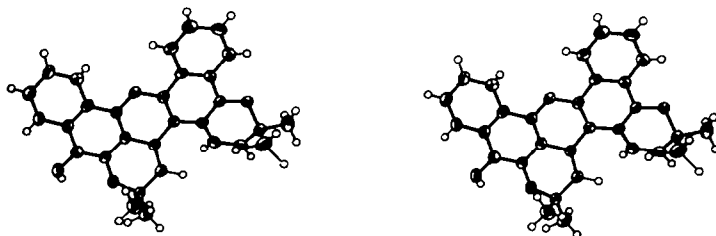
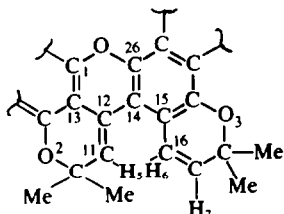


Fig. 2. The ORTEP X-ray crystallographic drawing of guayacanic acid.

The observed conformer appears to be dictated primarily by the steric interference between the two H atoms H(5) and H(6), shown in partial structure 4a.§ Inspection of molecular models shows that the distance between these two H atoms is a maximum when the axial Me groups are on the same side of the molecular plane. It is not apparent from models why the conformation illustrated in Fig. 2 is more stable than that with the two axial Me groups oriented away from the viewer. In the two alternative conformations, where the two axial Me groups are placed on opposite sides of the plane, the H(5)–H(6) distance is very small and, therefore, these conformations should be unstable.



4a (partial)

†In the text we have reported the reaction of naphthalene-1,4-diol with 11a and 12a to produce acetates 14 and 15 respectively. In one instance, however, this same reaction produced the opposite acetates (naphthalene-1,4-diol with 11a and 14a yielded 15 and 14 respectively). In this reaction, it is clear, that the same rearrangement observed in the synthesis of tetrahydroguayacanic acid (reaction of 12a with 13) has occurred. We have not, as yet, been able to duplicate this exceptional reaction.

‡Detailed X-ray data will be published elsewhere.

§The numbering system is that used in the X-ray investigation.

The H(5)–H(6) interaction leads to considerable stress in the molecule which results in a twist about the C(14)–C(15)–C(16) bond, so that the C(16) is forced considerably below the molecular plane as viewed in Fig. 2. C(11) is displaced 0.33 Å above the plane formed by C(1), C(12), C(13), C(14) and C(26). Both of these displacements increase the H(5)–H(6) distance, thus reducing the interaction between these two H atoms.

The easy synthesis of nondihydrolapachene, and tetrahydroguayacanic acid in aqueous media provides strong support for the hypothesis that 1,4 naphthaquinone (or

quinol) is an intermediate in the biosynthesis of naphthaquinone derivatives in higher plants.¹⁴ Furthermore, we have observed that prenyl naphthalene diols (11a, 12a, 13) undergo condensation even more readily than the unsubstituted naphthalene-1,4-diol itself, suggesting that these prenylated diols may be the intermediate biogenetic precursors of natural, dimeric prenylated naphthaquinone derivatives.

EXPERIMENTAL

All m.ps are uncorrected. NMR spectra were determined on a modified Varian HA-100 instrument in CDCl₃ with a TMS internal standard. Mass spectral analysis was determined on a CEC 110 high resolution mass spectrophotometer. IR data were obtained on a Perkin Elmer model 237B grating IR spectrophotometer. UV spectra were obtained on a Cary model 15 recording spectrophotometer.

Extraction. Hammermilled *Tabebuia guayacana* Hemsl. heartwood (5 kg) was successively hot solvent extracted with petroleum ether 30–60°, ethyl ether, acetone and methanol. Constituents from the ethyl ether and acetone extracts will be primarily considered in this investigation.

Ethyl ether extract

The hot ether extraction yielded 90 g of ether solubles and 35 g of insoluble residue upon cooling. The insoluble residue contained some lapachol. The ether solubles were concentrated to dryness, warmed with chloroform, and filtered.

Guayacanic acid (4a). The chloroform filtrate was cooled and the solid product was recrystallized from acetone-methanol to yield 4a as yellow cubic crystals, m.p. 225–226° (2.01 g; 0.04%). (Found: C, 79.7; H, 5.43. Calc. for C₂₀H₂₄O₄: C, 80.3; H, 5.39%); UV spectrum $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log ϵ), 405 (3.95), 385 (3.95), ~365 (3.86), ~295 (4.31), ~270 (4.67), 257 (4.76), ~251 (4.73), 238 (4.72), 218 (3.82). IR spectrum (Nujol): 3380 cm⁻¹, m; 1655 cm⁻¹, s; 1624 cm⁻¹, w; 1453 cm⁻¹, s; 1449 cm⁻¹, s; 1410 cm⁻¹, s; 1360 cm⁻¹, m;

1345 cm^{-1} , m; 1302 cm^{-1} , m; 1260 cm^{-1} , m; 1240 cm^{-1} , m; 1135 cm^{-1} , m; 1125 cm^{-1} , m; 1085 cm^{-1} , s; 1010 cm^{-1} , s; 910 cm^{-1} , m; 765 cm^{-1} , s. NMR spectrum: 6H, s, δ 1.56; 6H, s, δ 1.58; 1H, s, δ 5.60; 1H, d, J = 10 Hz, δ 5.73; 1H, d, J = 10 Hz, δ 7.05; 1H, s, δ 5.80; 4H, m, δ 7.42-7.70; 2H, m, δ 8.05-8.35; 2H, m, δ 8.36-8.65.

Guayacanic acetate (4b). Acetylation of **4a** ($\text{Ac}_2\text{O}/\text{pyr}$) produced **4b** as colorless needles (MeOH), m.p. 248-249°. (Found: C, 78.3; H, 5.42. Calc. for $\text{C}_{32}\text{H}_{30}\text{O}_5$: C, 78.35; H, 5.34%). NMR spectrum: 6H, s, δ 1.54; 6H, s, δ 1.59; 3H, s, δ 2.48; 1H, d, J = 10 Hz, δ 5.73; 1H, s, δ 5.83; 1H, d, J = 10 Hz, δ 7.02; 5H, m, δ 7.40-7.85; 1H, m, δ 8.18-8.30; 2H, m, δ 8.37-8.60.

O-Methyl guayacanic (4c). Methylation of **4a** (dimethyl sulfate/acetone) yielded **4c**, light yellow needles (MeOH), m.p. 205°. (Found: C, 80.3; H, 5.66. Calc. for $\text{C}_{31}\text{H}_{28}\text{O}_5$: C, 80.5; H, 5.67%). NMR spectrum: 12H, s, δ 1.58; 3H, s, δ 4.05; 1H, d, J = 10 Hz, δ 5.71; 1H, s, δ 5.82; 1H, d, J = 10 Hz, δ 7.03; 4H, m, δ 7.43-7.70; 2H, m, δ 8.00-8.30; 2H, m, δ 8.33-8.60.

Tetrahydroguayacanic 7a. Guayacanic **4a** (0.1 gm) was catalytically reduced in EtOAc, 5% Pd on C, 47 psi, 28 hr, with a two mole uptake of H_2 . The hydrogenated soln was filtered, concentrated and recrystallized (acetone/methanol) to yield **7a** as white plates, m.p. 249-250° (0.08 g). (Found: C, 79.5; H, 6.17. Calc. for $\text{C}_{30}\text{H}_{30}\text{O}_4$: C, 79.63; H, 6.24%). IR spectrum (Nujol): 3555 cm^{-1} , m; 1606 cm^{-1} , m; 1585 cm^{-1} , s; 1495 cm^{-1} , s; 1410 cm^{-1} , s; 1385 cm^{-1} , s; 1265 cm^{-1} , m; 1255 cm^{-1} , m; 1190 cm^{-1} , s; 1160 cm^{-1} , s; 1120 cm^{-1} , m; 1110 cm^{-1} , s; 1060 cm^{-1} , s; 980 cm^{-1} , m; 770 cm^{-1} , m; 760 cm^{-1} , NMR spectrum: 6H, s, δ 1.37; 3H, s, δ 1.53; 3H, s, δ 1.68; 4H, m, δ 1.60-2.03; 2H, m, δ 2.58-3.04; 1H, dd, J = 12 Hz, 3 Hz, δ 4.20; 1H, s, δ 5.52; 4H, m, δ 7.36-7.60; 2H, m, δ 8.00-8.26; 2H, m, δ 8.28-8.56.

Tetrahydroguayacanic acetate (7b). **7a** was acetylated ($\text{Ac}_2\text{O}/\text{pyr}$) to yield **7b** as colorless needles (MeOH), m.p. 244-245°. (Found: C, 77.6; H, 6.04. Calc. for $\text{C}_{32}\text{H}_{30}\text{O}_5$: C, 77.71; H, 6.11%). NMR spectrum: 3H, s, δ 1.35; 3H, s, δ 1.37; 3H, s, δ 1.54; 3H, s, δ 1.65; 3H, m, δ 1.72-2.05; 3H, s, δ 2.47; 3H, m, δ 2.60-3.05; 1H, dd, J = 11.5 Hz, 3 Hz, δ 4.21; 5H, m, δ 7.39-7.80; 1H, m, δ 8.13-8.30; 2H, m, δ 8.20-8.53.

O-Methyl-tetrahydroguayacanic (7c). **7a** was methylated (dimethyl sulfate/acetone) to yield **7c** as fine pale yellow needles (MeOH), m.p. 225-226°. (Found: C, 79.8; H, 6.47. Calc. for $\text{C}_{31}\text{H}_{28}\text{O}_4$: C, 79.8; H, 6.48%). NMR spectrum: 3H, s, δ 1.38; 3H, s, δ 1.43; 3H, s, δ 1.55; 3H, s, δ 1.71; 3H, m, δ 1.60-2.01; 3H, m, δ 2.60-3.03; 3H, s, δ 4.02; 1H, dd, J = 11.5 Hz, 3 Hz, δ 4.22; 4H, m, δ 7.40-7.65; 2H, m, δ 8.00-8.26, 2H, m, δ 8.33-8.54.

O-Methyl-dihydroguayacanic (9). **4a** (50 mg) in 80% aqueous formic acid (15 ml) was treated with 10% HCl aq (1 ml) and 1,3,6,8-tetrahydroxanthene (150 mg), and warmed until solid began to precipitate. Water was added and the mixture was extracted with ether. The ether soln was washed with water and sat. NaHCO_3 aq, dried and concentrated with MeOH to yield pink needles, m.p. 223-224°. (Found: C, 80.1; H, 6.23. Calc. for $\text{C}_{31}\text{H}_{28}\text{O}_4$: C, 80.15; H, 6.08%). NMR spectrum: (Fig. 1): 6H, s, δ 1.40; 3H, s, δ 1.62; 3H, s, δ 1.68; 1H, t, J = 13 Hz, δ 2.75; 1H, dd, J = 13 Hz, 3 Hz, δ 2.56; 3H, s, δ 3.99; 1H, dd, J = 13 Hz, 3 Hz, δ 4.20; 1H, d, J = 10 Hz, δ 5.67; 1H, d, J = 10 Hz, δ 6.47; 4H, m, δ 7.32-7.60; 2H, m, δ 7.98-8.22; 2H, m, δ 8.24-8.48.

Acetone extract

A portion of this extract was concentrated to dryness, and chloroform was added. The mixture was boiled, and filtered through celite. The soluble portion was evaporated to dryness, and MeOH was added. Tectol (**3**) crystallizes from the cooled MeOH soln as white granular crystals, m.p. 212° (d). (Found: C, 80.1; H, 5.79. Calc. for $\text{C}_{30}\text{H}_{28}\text{O}_4$: C, 79.98; H, 5.82%). NMR spectrum: 12H, s, δ 1.51; 2H, br, s, δ 5.06; 2H, d, J = 10 Hz, δ 5.55; 2H, d, J = 10 Hz, δ 5.96; 4H, m, δ 7.48-7.64; 4H, m, δ 8.15-8.22. Acetylation ($\text{Ac}_2\text{O}/\text{pyr}$) gave tectol diacetate as white needles (MeOH), m.p. 205-205°. Methylation (dimethylsulfate/acetone) gave tectol dimethyl ether as needles (MeOH), m.p. 216-217°. Observed physical and spectral properties, with the exception of NMR, of **3** and its derivatives are in close agreement with recorded values.⁴

Synthesis

Dehydro- α and β -lapachone diacetates (11b and 12b). The

diacetates **11b** and **12b** were synthesized from **1** (40 g) according to Hooker.¹¹ The synthesis yielded **dehydro- α -lapachone diacetate 11b** (19.9 g) as white needle clusters (MeOH), m.p. 131-132°. (Found: C, 70.0; H, 5.56. Calc. for $\text{C}_{19}\text{H}_{16}\text{O}_5$: C, 69.92; H, 5.65%); NMR spectrum: 6H, s, δ 1.42; 3H, s, δ 2.36; 3H, s, δ 2.38; 1H, d, J = 10 Hz, δ 5.82; 1H, d, J = 10 Hz, δ 6.46; 2H, m, δ 7.20-7.45; 2H, m, δ 7.53-7.78. The reaction also yielded **dehydro- β -lapachone diacetate 12b** (9.8 g) as colorless balls (MeOH), m.p. 129°. (Found: C, 70.0; H, 5.60. Calc. for $\text{C}_{19}\text{H}_{16}\text{O}_5$: C, 69.92; H, 5.65%); NMR spectrum: 6H, s, δ 1.48; 3H, s, δ 2.30; 3H, s, δ 2.37; 1H, d, J = 10 Hz, δ 5.68; 1H, d, J = 10 Hz, δ 6.33; 2H, m, δ 7.30-7.52, 2H, m, δ 7.53-7.72.

Nordihydrolapachenole (13). 2-Methyl-3-butene-2-ol was slowly added dropwise to a warm (85°) soln of naphthalene-1,4-diol (20 g) in 70% aq. formic acid (500 ml) containing ascorbic acid (5 g) (N_2). After 30 min, water was added, the mixture was cooled and the solid product was collected. A soln of this product in ether was diluted with MeOH and concentrated to yield a crystalline dichroman. The MeOH filtrate from the dichroman was evaporated to an oil. A soln of the oil in hexane gave, upon concentration and cooling, **13** as colorless needles, m.p. 126-128° (5.0 g; 18.8%) (M.S. observed M^+ : 228.1164. Calc. for $\text{C}_{17}\text{H}_{14}\text{O}_2$: 228.1150). NMR spectrum: 6H, s, δ 1.27; 2H, t, J = 5 Hz; 2H, t, J = 5 Hz; 1H, br, s, δ 5.03; 1H, br, s, δ 6.47; 2H, m, δ 7.30-7.52; 2H, m, δ 7.43-8.828.

Diacetate (14). A soln of **11b** (1.0 g) in a minimal amount of MeOH was treated with 10% NaOH aq (10 ml), containing sodium dithionite (3.0 g), warmed on a steam bath for 2 min, acidified with dil HCl aq and cooled. The solid product (**15a**) collected by gravity filtration (suction filtration accelerates oxidation) was washed with 5% aqueous ascorbic acid and used immediately without further purification. It was added to a soln of naphthalene-1,4-diol (0.80 g) in warm 80% aqueous formic acid (30 ml) and heated (stirring) for 2 hr. Excess of water was added and the mixture was extracted with ether. Evaporation of the dried ether extract gave a residual solid which was acetylated by warming with $\text{Ac}_2\text{O}/\text{pyridine}$. Addition of water, extraction with ether, drying and concentration with MeOH yielded colorless granular crystals **14**, m.p. 276-277°. (Found: C, 74.0; H, 4.93. Calc. for $\text{C}_{29}\text{H}_{22}\text{O}_6$: C, 74.67; H, 4.75%); NMR spectrum: 6H, s, δ 1.55; 6H, s, δ 2.47; 1H, s, δ 5.79; 1H, s, δ 7.42; 6H, m, δ 7.40-7.85; 2H, m, δ 8.30-8.60.

Diacetate (15). **12b** was deacetylated as for **11b**, the solid product (**12a**) was immediately added to a soln of naphthalene-1,4-diol in 80% aqueous formic acid, heated for 2 hr, water added, extracted with ether, and acetylated. **15** was crystallized (acetone/methanol) as yellow needles, m.p. 249-250°. (Found: C, 74.6; H, 4.88. Calc. for $\text{C}_{29}\text{H}_{22}\text{O}_6$: C, 74.67; H, 4.75%); NMR spectrum: 6H, s, δ 1.63; 3H, s, δ 2.47; 3H, s, δ 2.62; 1H, s, δ 5.81; 1H, s, δ 7.45; 6H, m, δ 7.46-7.90; 2H, m, δ 8.10-8.35.

Tetrahydroguayacanic (12a). **11a** (2 gm) was immediately added to **13** (2 g) in 80% aq. formic acid containing ascorbic acid (5 g) (N_2), reacted for 2 hr, the mixture poured into water, cooled overnight and the solid filtered off and washed with water. The solid was dissolved in acetone, concentrated and crystallized (MeOH) as colorless granular crystals m.p. 251-253° (1.7 g; 43%). (Found: C, 79.48; H, 6.2. Calc. for $\text{C}_{30}\text{H}_{28}\text{O}_4$: C, 79.62; H, 6.24%). The physical and spectral properties (m.p., NMR, IR, UV) were identical to the tetrahydroguayacanic obtained in the catalytic reduction of guayacanic.

Attempted synthesis of 8a. **12a** was reacted with **13** exactly as in the previously described reaction (**11a** and **13**). The reaction was worked up to yield granular crystals (acetone/methanol) m.p. 251-253° (8% yield). (Found: C, 79.5; H, 6.27. Calc. for $\text{C}_{30}\text{H}_{28}\text{O}_4$: C, 79.62; H, 6.24%). The physical and spectral properties were identical to synthetic and natural **7a**. This product was therefore designated **7a** rather than **8a**.

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REFERENCES

- C. R. Southwell and J. D. Bultman, *Biotropica* **3**(1), 81 (1971).
- L. E. Wise, R. C. Rittenhouse and C. Garcia, *Tappi* **34**(4), 185 (1951).

- ³A. R. Burnett and R. H. Thomson, *J. Chem. Soc. (C)*, 2100 (1967).
⁴A. R. Burnett and R. H. Thomson, *Ibid. (C)*, 850 (1968).
⁵W. Sandermann and H. H. Dietrichs, *Holz als Roh- und Werkstoff* **15**, 281 (1957).
⁶W. Sandermann and M. H. Simatupang, *Ibid.* **24**, 190 (1966).
⁷H. H. Dietrichs, *Naturwiss.* **51**, 408 (1964).
⁸W. Sanderman and H. H. Dietrichs, *Holzforschung* **13**, 137 (1959).
⁹L. Jurd and T. C. Somers, *Phytochem.* **9**, 419 (1970).
¹⁰W. F. Kröhnke and K. Dickoré, *Chem. Ber.* **92**, 46 (1959).
¹¹S. C. Hooker, *J. Am. Chem. Soc.* **58**, 1190 (1936).
¹²M. G. Ettlinger, *Ibid.* **72**, 3090 (1950).
¹³C. K. Johnson, *ORTEP Rep.* ORNL-3794, Oak Ridge National Laboratory, Oak Ridge, Tennessee.
¹⁴R. H. Thomson, *Naturally Occurring Quinones*, p. 20. Academic Press, New York (1971).